



**MORGAN**<sup>TM</sup>  
PATUXENT ENVIRONMENTAL AND  
AQUATIC RESEARCH LABORATORY

# *From Spat to Spectacular:*

Egg Quality Assessment and Probiotic  
Intervention in Oyster Larvae

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# Overview

01

## Background

Introduction to  
Oyster aquaculture  
and challenges in  
hatchery  
production

02

## Methods

Methods used to  
complete the study

03

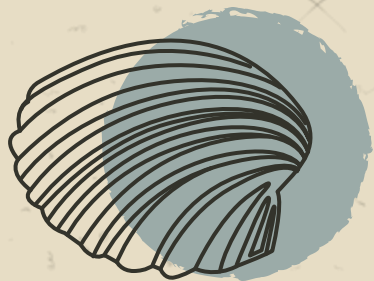
## Results

% Survivorship  
Growth

04

## Discussion

Conclusions and  
Key takeaways



*01*

# *Background Information*

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## *Oyster Aquaculture in the US*

**1600s–1800s:** Wild oyster harvesting was common along the East Coast

**1840s:** First commercial oyster farms established in New York

**Late 1800s:** Oyster Industry boomed, especially in the Chesapeake Bay

**Early 1900s:** Overharvesting and pollution led to a decline in wild oyster populations

**1920s–1950s:** Development of hatchery techniques and off-bottom culture methods

# *Oyster Aquaculture in the Chesapeake*

**1960s:** Dramatic decline in wild oyster populations due to overharvesting and diseases

**1980s:** Introduction of aquaculture to supplement wild harvest

**2000s:** Streamlining of permitting process to encourage oyster farming

**2010s:** Significant growth in oyster aquaculture production



# *Hatchery Seed Production*



## *Importance*

- Provides consistent and reliable source of larvae
- Allows for selective breeding and genetic improvements
- Enables year-round production
- Production of triploid (sterile) oysters for faster growth



## *Process and Benefits*

- Controlled spawning of broodstock
- Larval rearing in optimal conditions
  - Setting of larvae on micro cultch
- Nursery phase for juvenile growth
  - Increased survival compared to wild-caught seed
- Ability to produce specific strains

# *Challenges in hatchery production*



## *Water Quality*

Fluctuations in dissolved oxygen, pH, and nutrients

Presence of contaminants or pollutants



## *Environmental Conditions*

Temperature control (climate change)

Salinity management for optimal growth



## *Food Quality*

Maintaining algal cultures

Ensuring nutritional adequacy for different life stages

# *Biotic Challenges*



## *Microbial Community*

Balancing beneficial and harmful bacteria  
Controlling biofilm in rearing systems



## *Pathogen Control*

Prevention and management of disease

## *Egg Quality*

Maintaining healthy and diverse broodstock  
Optimizing conditioning and spawning protocols



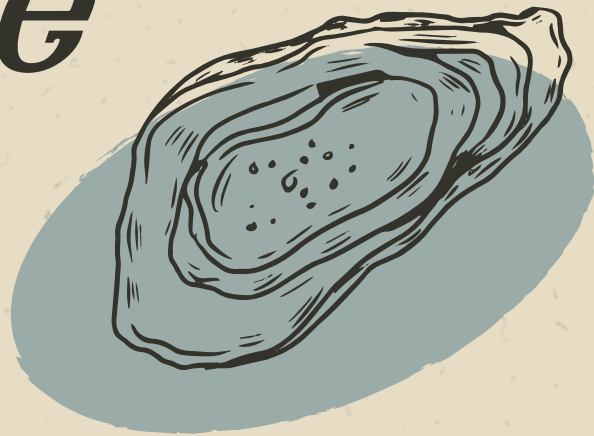
## *Other*

Managing predators or competitors in nursery systems  
Optimizing larval settlement





# *Goals of the Study*



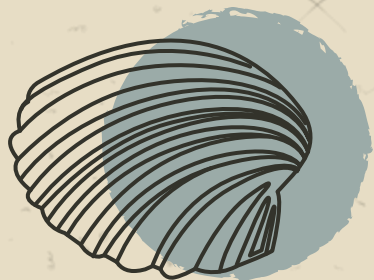
# *Investigating two factors that impact breeding SUCCESS*

1) Broodstock female Egg  
quality, including egg grade,  
and egg numbers

2) If adding probiotics can  
enhance larval yield or larval  
growth

02

# *Methods*





## *Strip Spawning*

Oyster 292 (Randy)



# *Objective 1 - egg quality*



## *Strip Spawning Broodstock Oysters*

80 oysters were shucked and their gametes checked for health and sex



## *Grading Eggs*

Females are given a number (1-3) for their egg quantity and their egg quality

Only the top scores will be selected

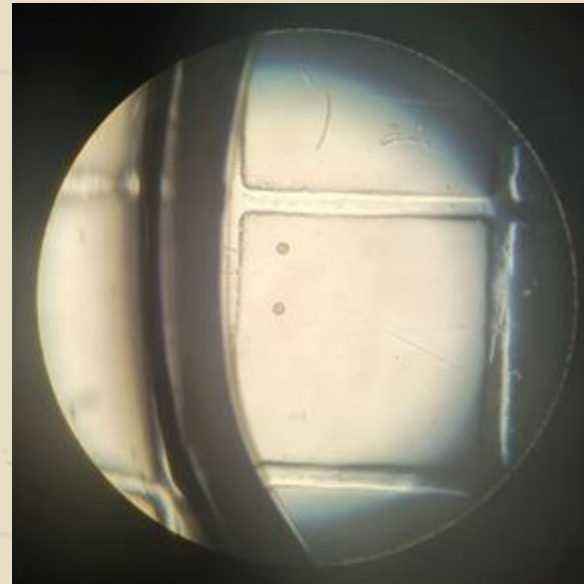
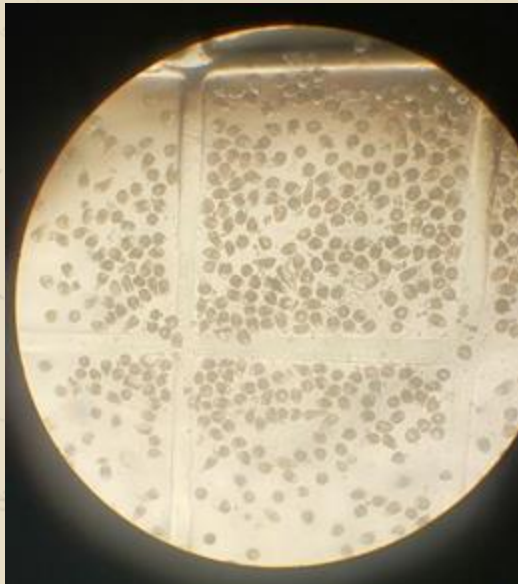


## *Final Groupings*

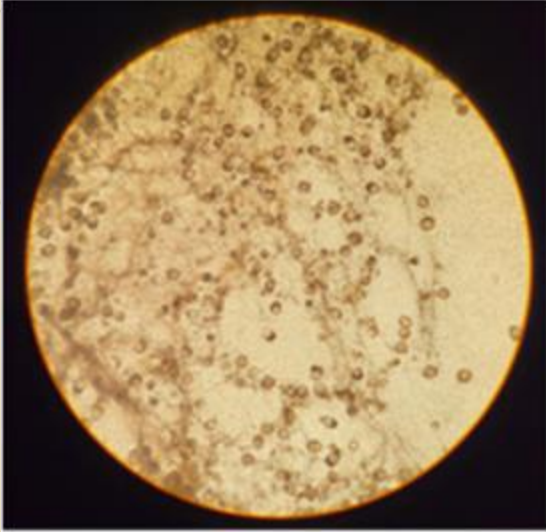
12 Females and 12 Males were selected for this study

## *Good Quantity vs. Good Quality*

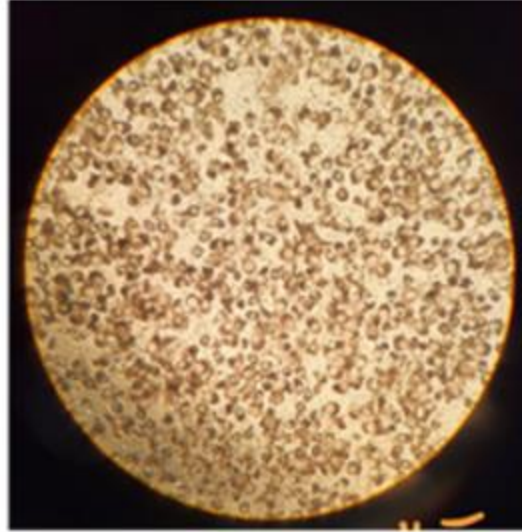
1) Female 28 (25.5M) – Good Quantity    2) Female 63 (48K) – Good Quality



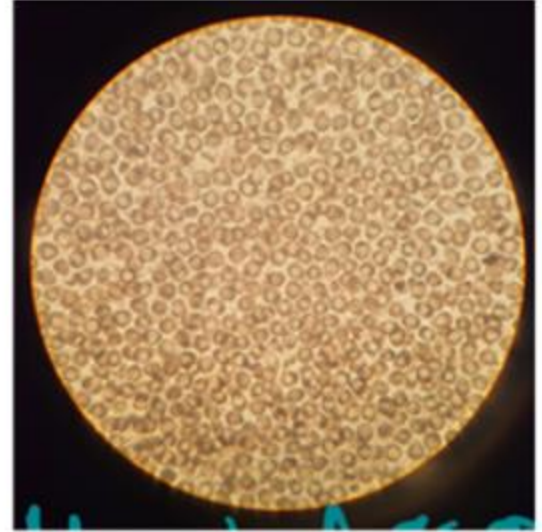
## *Egg Quality Grading cont.*



Grade 1



Grade 2



Grade 3

## *Creating Lines*

Female #	Egg Quantity	Egg Quality	Group #
28	25.53 M	1.5	1
31	109 K	2.5	2
38	413 K	2.5	2
41	183 K	2	2
47	760 K	2.5	2
51	121 K	2	2
53	50 K	2	2
63	48 K	3	2
76	82 K	1	3
78	2500	1	3
23	2.49 M	1	3
59	170 K	1	3

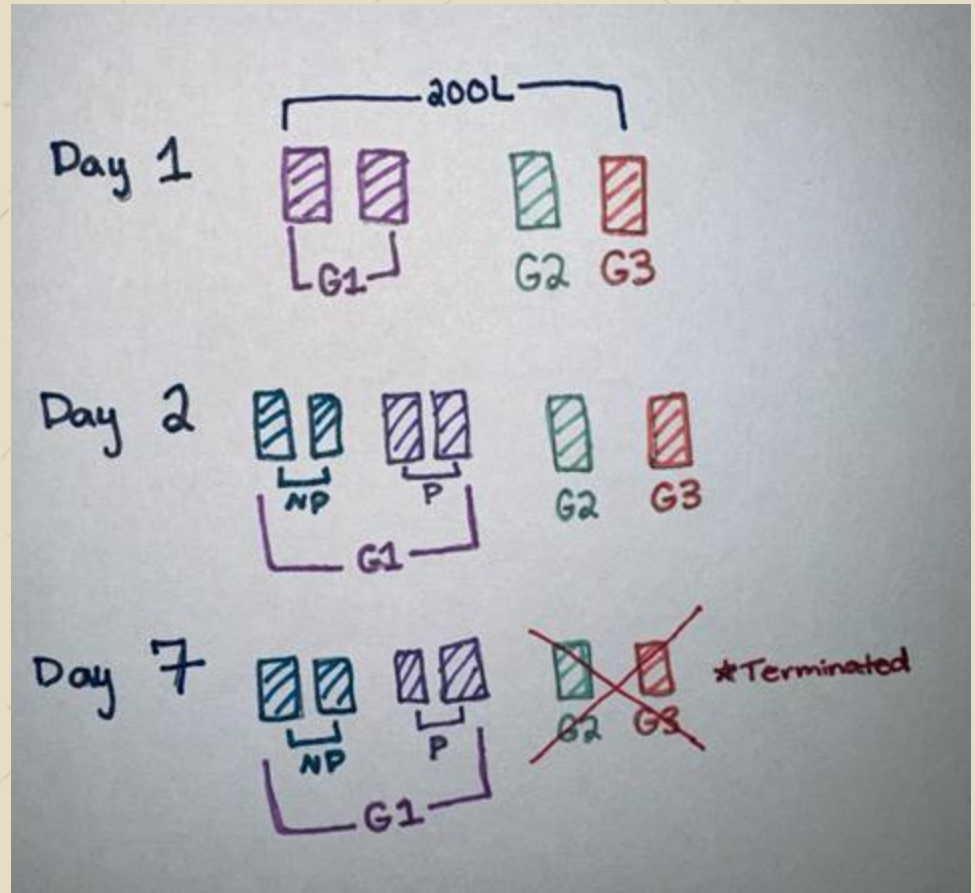
Group 1 – one female, highest egg quantity

Group 2 – seven females, highest quality

Group 3 – four females, lowest quality



# Experimental Design



# Objective 2 - Probiotic intervention



Control vs. Replicate Groups





## *Probiotic Utilized*

100 ml added after water change

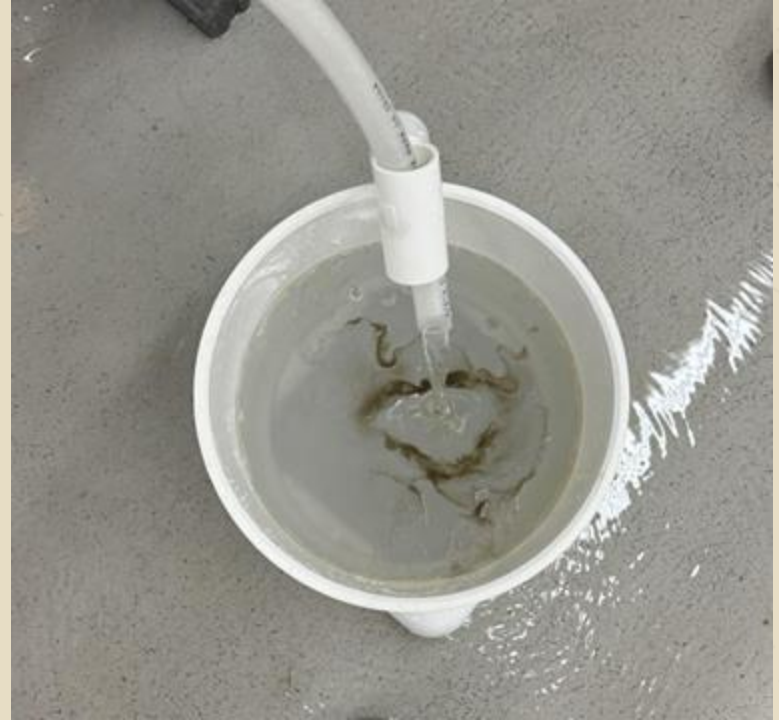
Hatchery Prime Tablets Claims:

- Reduces vibrio populations
- Eliminates toxic ammonia
- Increases overall yield
- Enhances water quality

“Using Hatchery Prime results in an 11.12% increase in survival”

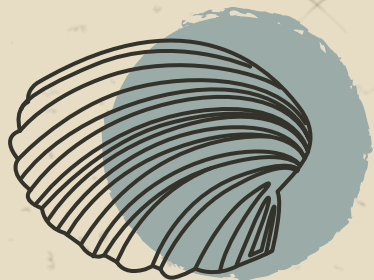
## *Overall Methods*

- Larvae Fed daily
- Larval Drops – MWF
  - Siphon larvae onto corresponding sieves
  - Counting for density
  - Size them for feeding

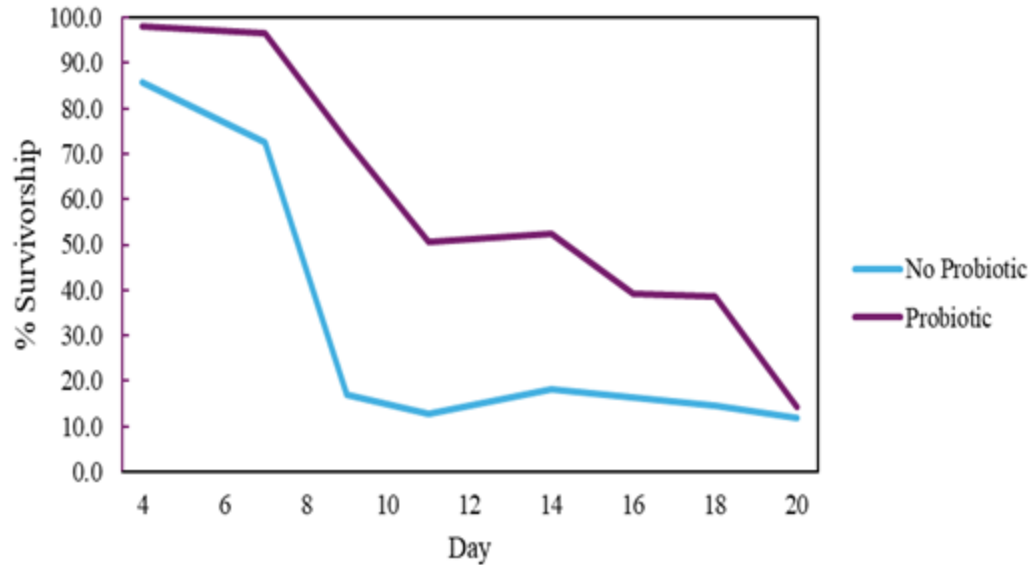


03

# *Results*



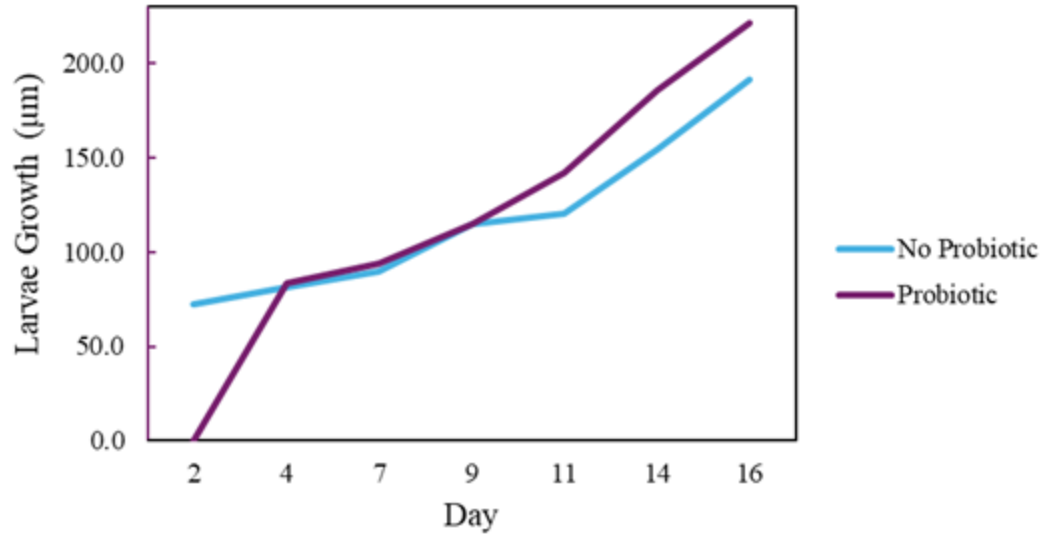
# Percent Survival

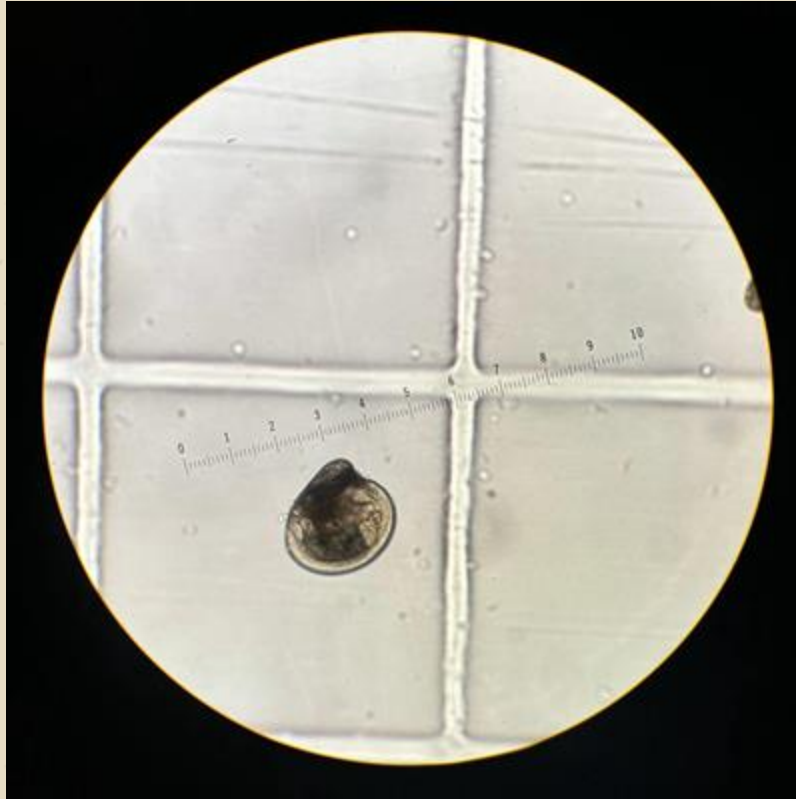


- Initial Survival both groups: high
- Probiotic group maintains higher survival throughout the entire study
- Day 20: probiotic group has a survival rate ~1.5x higher than the non-probiotic group

# *Growth Rate*

- Probiotic group starts lower but catches up by day 4
- Day 11, probiotic group shows faster growth
- Able to harvest probiotic group at day 16





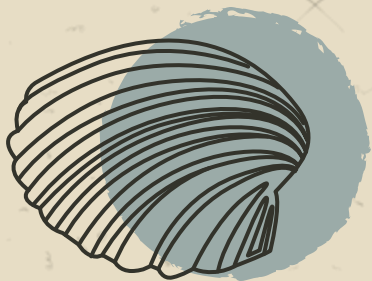
*Harvest Size*





04

# *Discussion*





## *Conclusion*

### *Addressing Larval Crashes*

Reduce crashes and  
increase larval survival to  
setting stage

Importance of shortening  
days to harvest

### *Probiotic Effects*

Survival: increased  
survivorship

Growth: accelerated  
development in probiotic  
group

# *Egg Quantity vs. Egg Quality*

Egg Quantity over Egg Quality in this study (go female 28!)

Group 2 and 3 did not make it to harvest

Future work required to truly determine those results

Only one female scored a 3

Broodstock – naturally conditioned

- Did not control feed amounts or temperature
- Prime spawning is at the end of May and early June
- Experiment ran through the end of July

# *Environmental Adaptations*

Oyster larvae struggle below 10  
ppt salinity

Chesapeake Bay is having a low  
salinity year

Solution: Added salt to water  
(12ppt)

Result: increased oyster  
production in low salinity  
conditions



## *Key Takeaways and Future Directions*



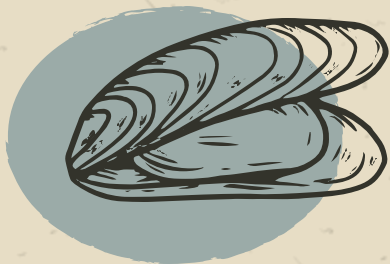
1. Probiotics show promise for improving survival and growth
1. Egg quantity over quality in this case
1. Potential for salinity management in oyster aquaculture

*35,377*

Individual larvae counted

*124*

White Boards cleared



*Tetraselmis*

Favorite Algae

## *Citations*

Utting, S. D., & Millican, P. F. (1997). Techniques for the hatchery conditioning of bivalve broodstocks and the subsequent effect on egg quality and larval viability. *Aquaculture*, 155(1-4), 45-54.

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*THANK YOU!*

