



MORGAN
PATUXENT ENVIRONMENTAL AND
AQUATIC RESEARCH LABORATORY

From Spat to Spectacular:

Egg Quality Assessment and Probiotic
Intervention in Oyster Larvae

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Overview

01

Background

Introduction to
Oyster aquaculture
and challenges in
hatchery
production

02

Methods

Methods used to
complete the study

03

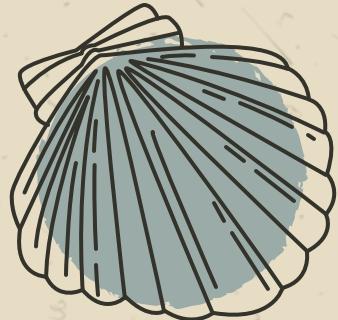
Results

% Survivorship
Growth

04

Discussion

Conclusions and
Key takeaways



Background Information

01

Oyster Aquaculture in the US



1600s–1800s: Wild oyster harvesting was common along the East Coast

1840s: First commercial oyster farms established in New York

Late 1800s: Oyster Industry boomed, especially in the Chesapeake Bay

Early 1900s: Overharvesting and pollution led to a decline in wild oyster populations

1920s–1950s: Development of hatchery techniques and off-bottom culture methods

Oyster Aquaculture in the Chesapeake

1960s: Dramatic decline in wild oyster populations due to overharvesting and diseases

1980s: Introduction of aquaculture to supplement wild harvest

2000s: Streamlining of permitting process to encourage oyster farming

2010s: Significant growth in oyster aquaculture production



Hatchery Seed Production



Importance

- Provides consistent and reliable source of larvae
- Allows for selective breeding and genetic improvements
- Enables year-round production
- Production of triploid (sterile) oysters for faster growth



Process and Benefits

- Controlled spawning of broodstock
- Larval rearing in optimal conditions
- Setting of larvae on micro cultch
- Nursery phase for juvenile growth
- Increased survival compared to wild-caught seed
- Ability to produce specific strains

Challenges in hatchery production



Water Quality

Fluctuations in dissolved oxygen, pH, and nutrients

Presence of contaminants or pollutants



Environmental Conditions

Temperature control (climate change)

Salinity management for optimal growth



Food Quality

Maintaining algal cultures

Ensuring nutritional adequacy for different life stages

Biotic Challenges

Microbial Community



Balancing beneficial and harmful bacteria

Controlling biofilm in rearing systems

Pathogen Control



Prevention and management of disease

Egg Quality

Maintaining healthy and diverse broodstock

Optimizing conditioning and spawning protocols



Other

Managing predators or competitors in nursery systems

Optimizing larval settlement

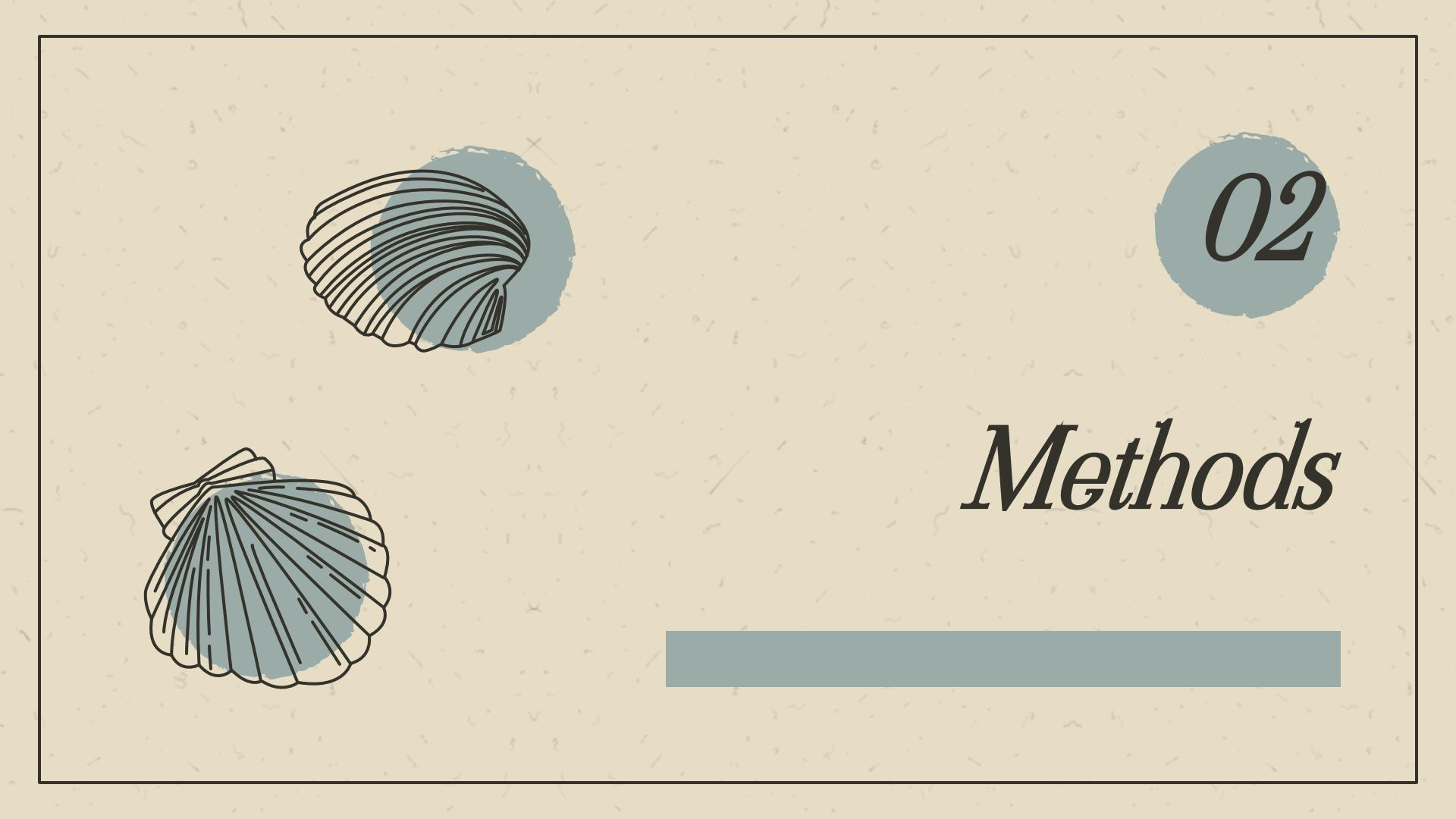


Goals of the Study



Investigating two factors that impact breeding success

- 1) Broodstock female Egg quality, including egg grade, and egg numbers
- 2) If adding probiotics can enhance larval yield or larval growth



02

Methods





Strip Spawning

Oyster 292 (Randy)

Objective 1 - egg quality



Strip Spawning Broodstock Oysters

80 oysters were shucked and their gametes checked for health and sex



Grading Eggs

Females are given a number (1-3) for their egg quantity and their egg quality

Only the top scores will be selected

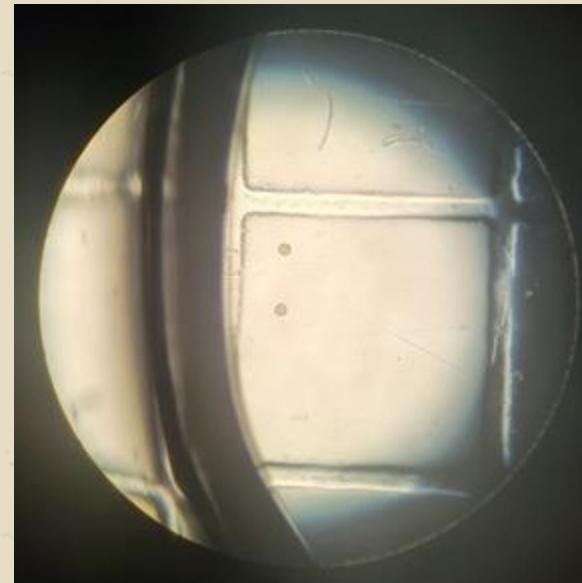
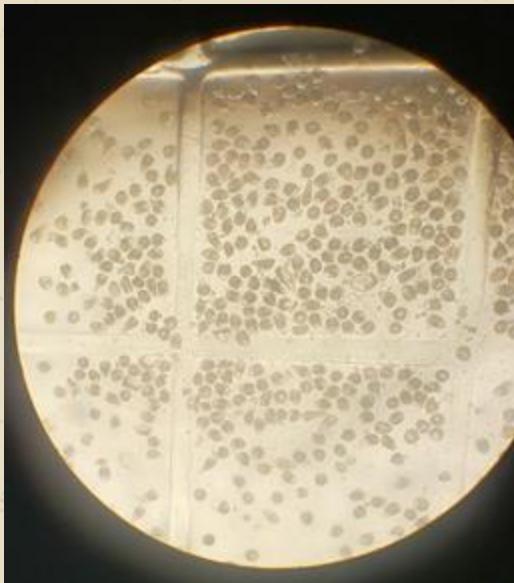


Final Groupings

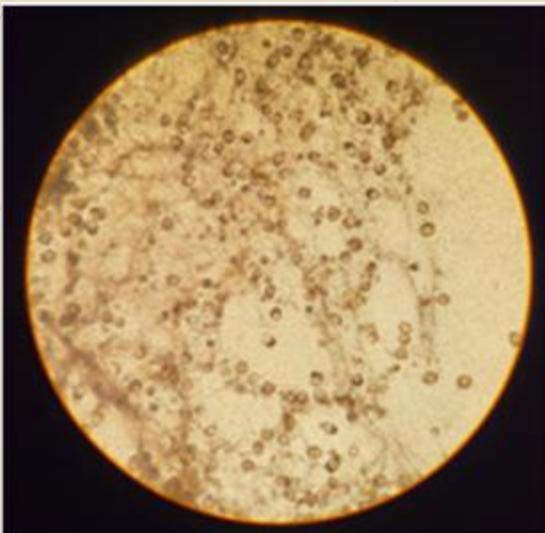
12 Females and 12 Males were selected for this study

Good Quantity vs. Good Quality

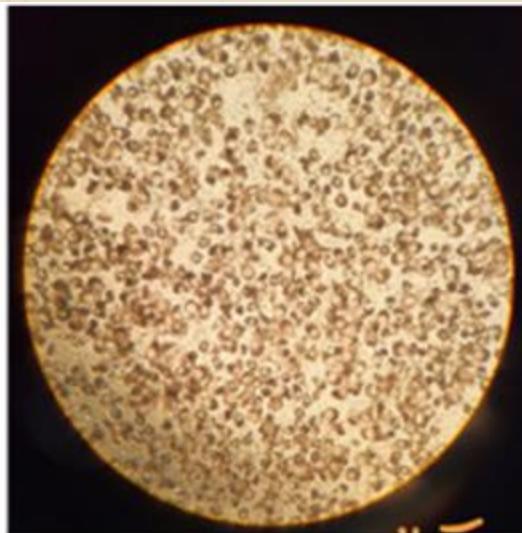
1) Female 28 (25.5M) - Good Quantity 2) Female 63 (48K) - Good Quality



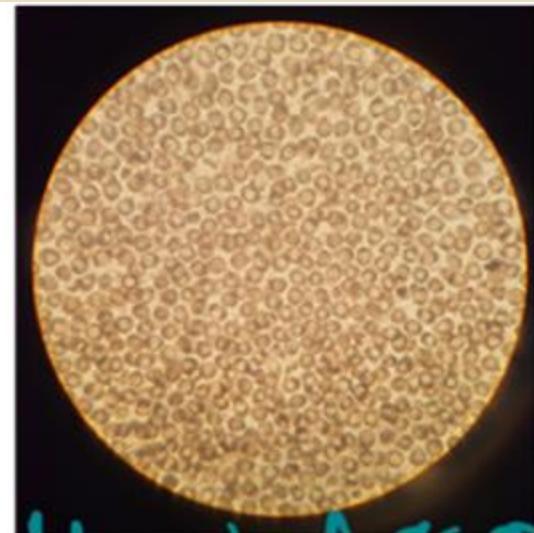
Egg Quality Grading cont.



Grade 1



Grade 2



Grade 3

Creating Lines

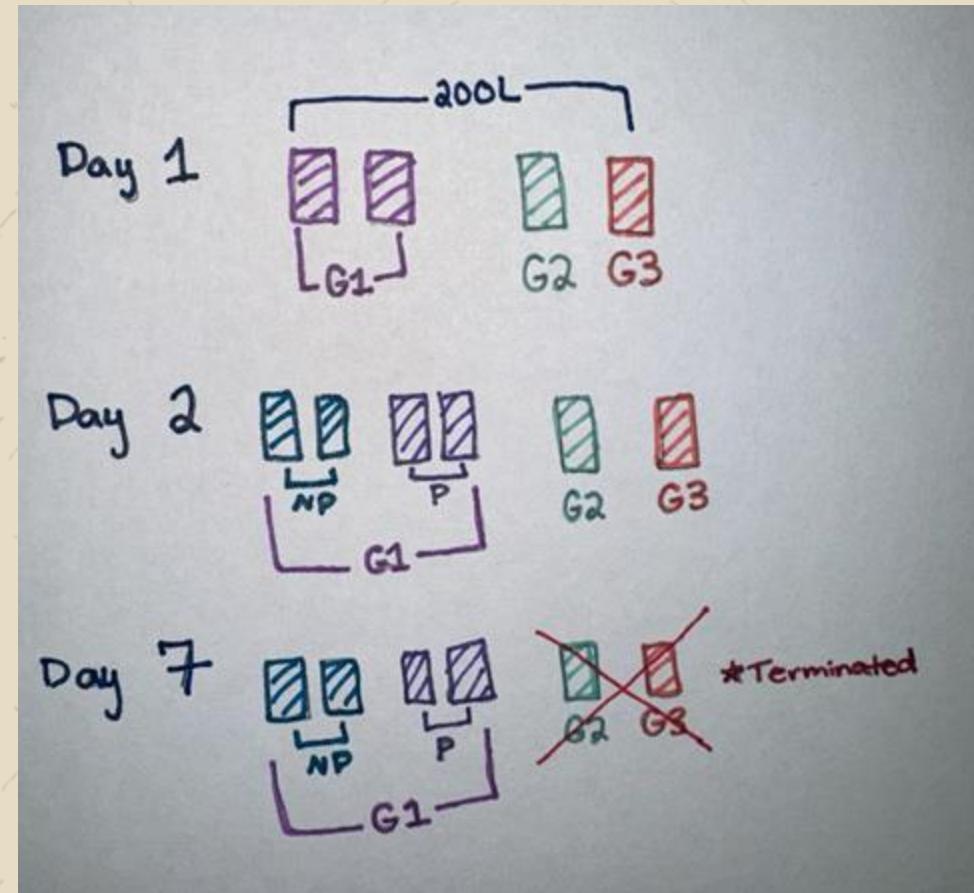
Female #	Egg Quantity	Egg Quality	Group #
28	25.53 M	1.5	1
31	109 K	2.5	2
38	413 K	2.5	2
41	183 K	2	2
47	760 K	2.5	2
51	121 K	2	2
53	50 K	2	2
63	48 K	3	2
76	82 K	1	3
78	2500	1	3
23	2.49 M	1	3
59	170 K	1	3

Group 1 – one female, highest egg quantity

Group 2 – seven females, highest quality

Group 3 – four females, lowest quality

Experimental Design



Objective 2 - Probiotic intervention



Control vs. Replicate Groups



Probiotic Utilized



100 ml added after water change
Hatchery Prime Tablets Claims:

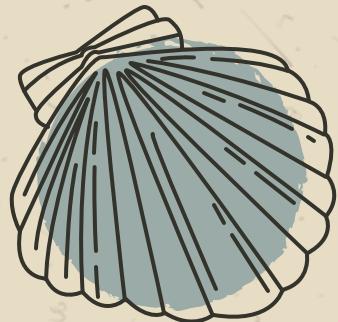
- Reduces vibrio populations
- Eliminates toxic ammonia
- Increases overall yield
- Enhances water quality

“Using Hatchery Prime results in an 11.12% increase in survival”

Overall Methods

- Larvae Fed daily
- Larval Drops – MWF
 - Siphon larvae onto corresponding sieves
 - Counting for density
 - Size them for feeding

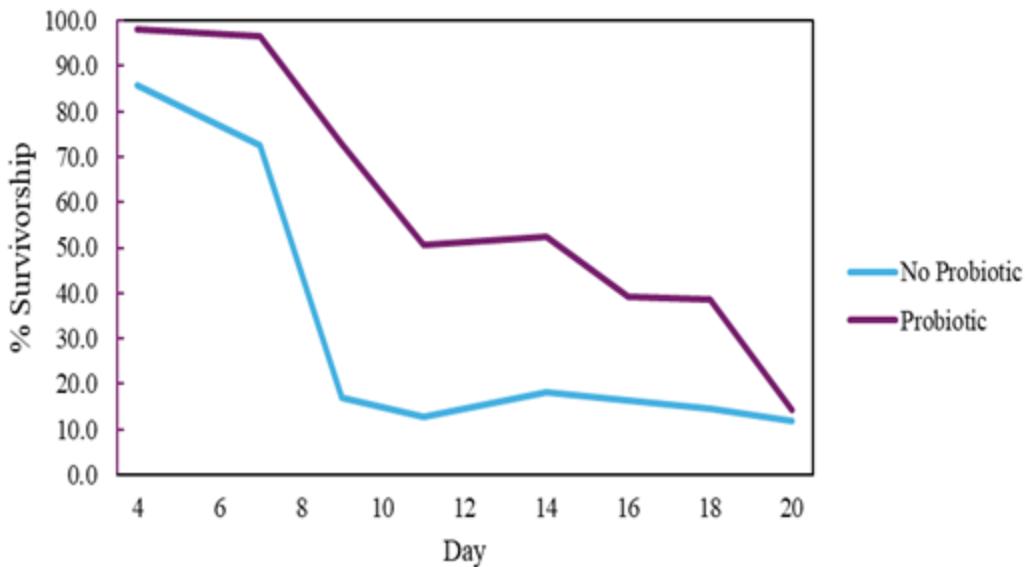




03

Results

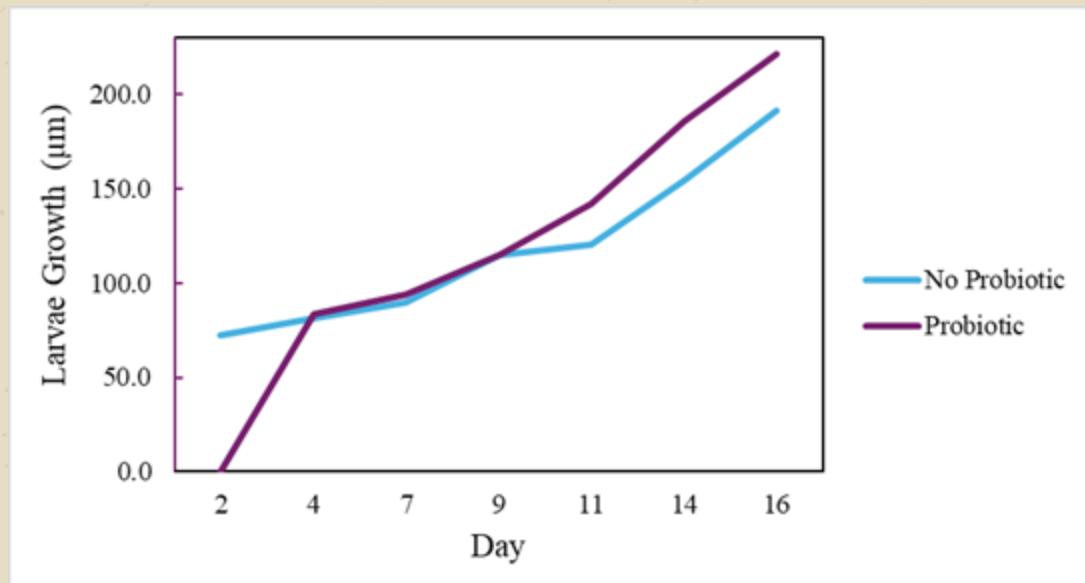
Percent Survival

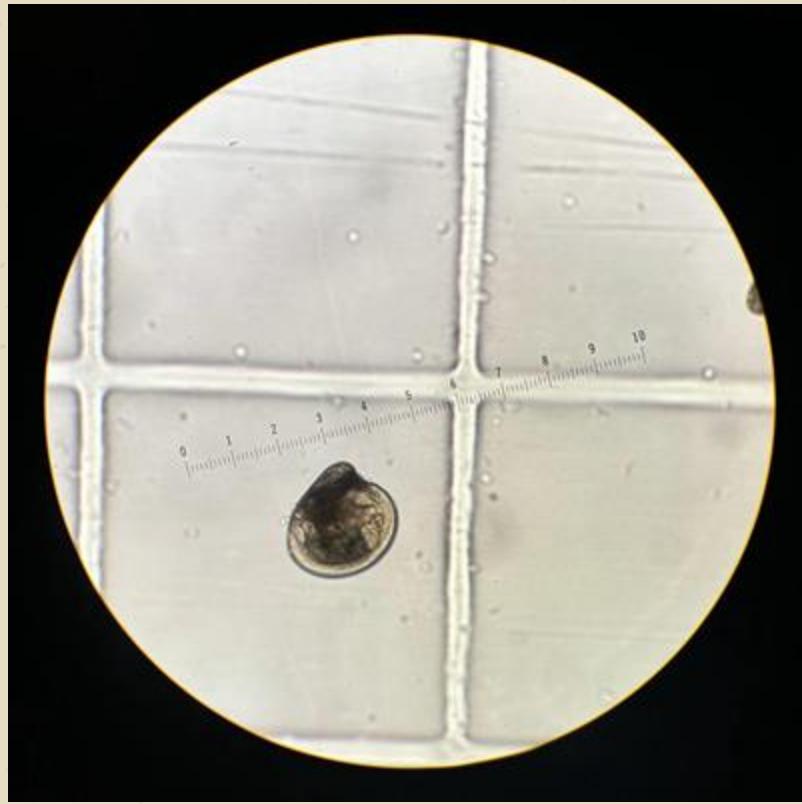


- Initial Survival both groups: high
- Probiotic group maintains higher survival throughout the entire study
- Day 20: probiotic group has a survival rate ~1.5x higher than the non-probiotic group

Growth Rate

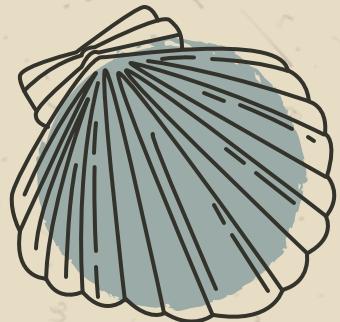
- Probiotic group starts lower but catches up by day 4
- Day 11, probiotic group shows faster growth
- Able to harvest probiotic group at day 16





Harvest Size





Discussion

04



Conclusion

Addressing Larval Crashes

Reduce crashes and increase larval survival to setting stage

Importance of shortening days to harvest

Probiotic Effects

Survival: increased survivorship

Growth: accelerated development in probiotic group

Egg Quantity vs. Egg Quality

Egg Quantity over Egg Quality in this study (go female 28!)

Group 2 and 3 did not make it to harvest

Future work required to truly determine those results

Only one female scored a 3

Broodstock – naturally conditioned

- Did not control feed amounts or temperature
- Prime spawning is at the end of May and early June
- Experiment ran through the end of July

Environmental Adaptations

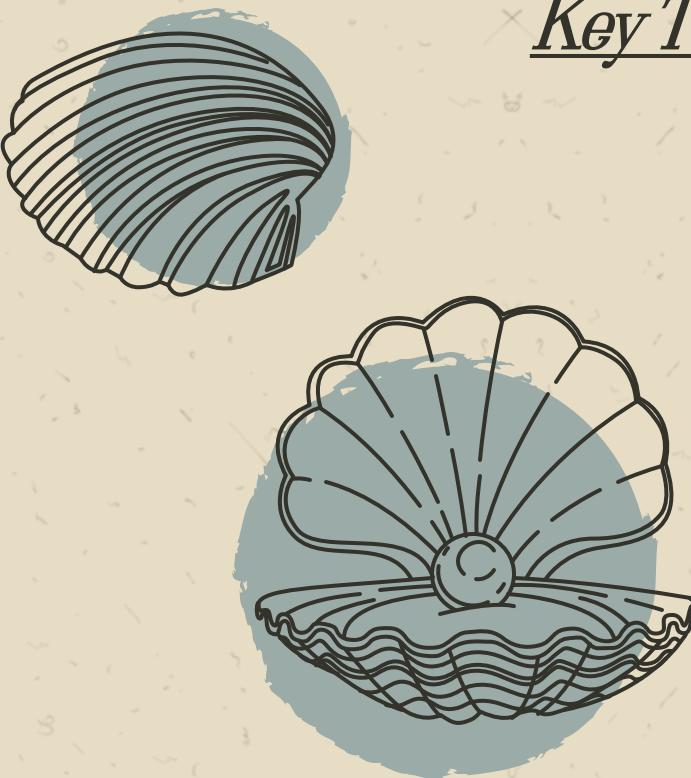
Oyster larvae struggle below 10
ppt salinity

Chesapeake Bay is having a low
salinity year

Solution: Added salt to water
(12ppt)

Result: increased oyster
production in low salinity
conditions





Key Takeaways and Future Directions

1. Probiotics show promise for improving survival and growth
1. Egg quantity over quality in this case
1. Potential for salinity management in oyster aquaculture

35,377

Individual larvae counted

124

White Boards cleared



Tetraselmis

Favorite Algae

Citations

Uutting, S. D., & Millican, P. F. (1997). Techniques for the hatchery conditioning of bivalve broodstocks and the subsequent effect on egg quality and larval viability. *Aquaculture*, 155(1-4), 45-54.

Gallagher, SM.. Mann. R.. 1986. Growth and survival of larvae of Mercman. *CL.1 and Crusso.strm virginica* relative to broodstock conditioning and lipid content of eggs. *Aquaculture* 56, 105-121.

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Acknowledgments

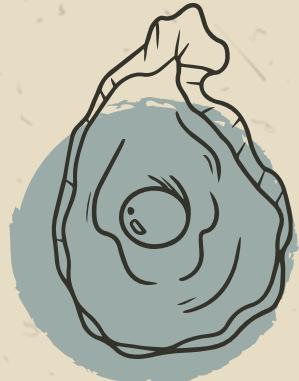


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Congressionally Directed Spending 2023 (Earmark) "Morgan State University's PEARL Lab Student Research Enhancements."

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Emotional Support Tape Ball





THANK YOU!

